

**Steroidal Sapogenin from the Rhizomes of *Asparagus africanus* Lam**

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A steroidal sapogenin, yamogenin (25S-spirost-5-en-3 $\beta$ -ol) was isolated from the petroleum ether extract of the rhizomes of *Asparagus africanus*. The structure of the sapogenin was elucidated by spectral analysis.

**Key word:** *Asparagus africanus*, rhizomes, steroidal sapogenin, yamogenin

**INTRODUCTION**

Plants of the family Liliaceae are known to be rich sources of steroidal saponins [1]. The genus *Asparagus* is widely distributed in tropical and temperate zone [2]. Some *Asparagus* species have already been studied and found to contain steroidal saponins [3]. In different parts of Africa, *Asparagus africanus* is used in traditional medicine as diuretics and haematuric agents, other uses include as analgesia in rheumatoid arthritis, chronic gout, and haemorrhoid [4, 5,6]. A steroidal sapogenin known as muzanzagenin [12 $\beta$ , 12 $\alpha$ -di hydroxy-(25R)-spirosta -4,7-dien-3-one) has been isolated from the root of *A. Africanus* [7]. This paper reports the isolation of yamogenin [1] from the rhizomes of *A Africanus*.

**MATERIALS AND METHODS**

Melting point was determined using electrothermal-melting point apparatus and reported uncorrected. Column chromatography was carried out on silica gel G (60-120 $\mu$ m) BDH product, TLC on silica gel G. BDH product, I.R was recorded on Pye Unicam series fourier transform spectrophotometer. The HPLC-APCI-MS was carried out on Esquire LC 00028 HPLC-MS. EIMS was carried out on HSE 8964 spectrophotometer 70EV.

**Plant Material:** The rhizomes of *A. africanus* were collected in a village near Samaru, Zaria-Nigeria, and identified by comparison with a specimen (vouchered No 900129) deposited at the herbarium of the Department of Biological Sciences, Ahmadu Bello University, Zaria.

**Extraction:** The air-dried powdered rhizomes of the plant (300g) was exhaustively extracted with petroleum ether (60-80 $^{\circ}$ C) with a soxhlet extractor for 24hr. The petroleum ether extract was concentrated at reduced pressure to give a yellow residue (4.55g). part of which (3g) was subjected to extensive column chromatography over silica gel and eluted gradiently with petroleum ether (60-80 $^{\circ}$ C) through chloroform and methanol. Fractions (40-80) pooled together from petroleum ether: chloroform (80:20) afforded a major compound that was crystallized in methanol to give a white crystals (35mg) which has a melting point of 195-197 $^{\circ}$ C. TLC of the compound gave RF values of 0.6, 0.67 and 0.2 using petroleum ether: Acetone (80:20); petroleum ether: methanol (80:20) and chloroform: petroleum ether (80:20) respectively.

**RESULT**

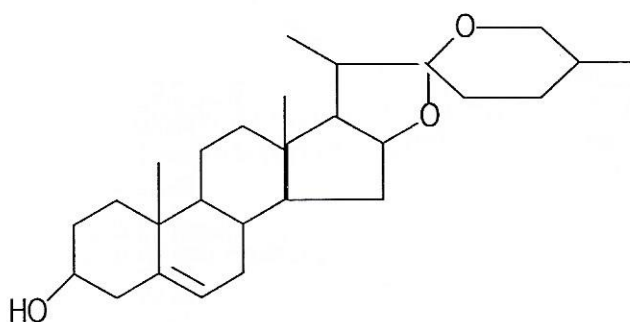
**Spectral Properties:** IR (Nujol) cm $^{-1}$ , 3523 (OH), 1654 (C=C), 1040 (C-O), 963, 919, 900, 847 (intensity 919 > 900, 25S-spirostanol).

HPLC-APCI-MS: [M+H] $^{+}$ 415.

EIMS (PROBE). 70ev M/z (relative intensity): 414 (M $^{+}$ ) (8), 342 (22), 300 (34), 282 (65), 271 (30), 139 (100), 115 (15).

**DISCUSSION**

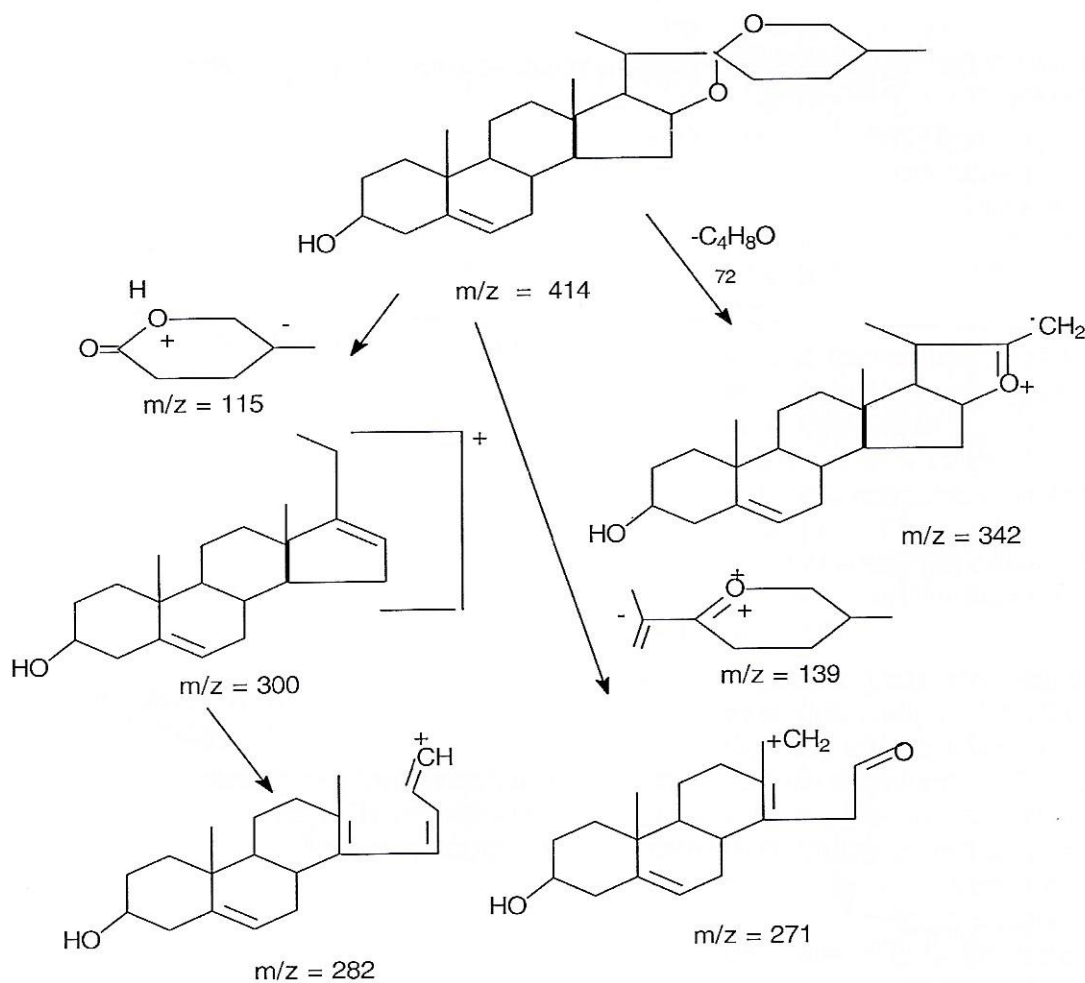
The petroleum ether extract of the rhizomes of *A. africanus* after chromatography separation gave a compound characterized as yamogenin. The IR spectrum shows bands for OH (3523cm $^{-1}$ ), C=C (1654cm $^{-1}$ ), C-O (1040cm $^{-1}$ ), and the spirostan structure (963,919,900,840cm $^{-1}$ ) with 919cm $^{-1}$  stronger than 900cm $^{-1}$  which shows a 25S-sapogenin [8].



Yamogenin  
(25S-Spirost-5en-3β-01)

The HPLC – APCI-MS showed molecular weight to be 414 which corresponds to molecular formula:  $C_{27}H_{42}O_3$ , this point to an epimer of Diosgenin. Figure 1 shows typical characteristics mass spectra fragments formed by the cleavage of E and F rings of this compound [9, 10]. The EI-MS spectrum of

an authentic sample of diosgenin was compared with that of the compound and found to contain similar fragmentation pattern. Co-TLC of the compound with authentic diosgenin using solvent system: chloroform: methanol (95:5) gave same RF value (0.58).



**Figure 1: Mass fragmentation pattern of yamogenin ( $C_{27}H_{42}O_3$ )**

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#### REFERENCES

1. Bernardo, R.R., Pinto, A.V. and Parente, J.P. (1996). Steroidal Saponins from *Smilax officinalis*. *Phytochemistry*, **43**(2), 465-469.
2. Evans, J.M. (1956). Trease and Evans' Pharmacognosy. 14<sup>th</sup> edition. Published by W.B. Saunders Company Ltd. U.K. pp20-50.
3. Mahato, S.B., Ganguly, A.N. and Sahu, N.P. (1982). Steroidal Saponin. A review. *Phytochemistry*, **21** (5), 959-978.
4. Dalziel, J.M (1956). The Useful Plants of West Tropical Africa. Crown Agents for Overseas Government and Administration, London Pp. 477.
5. Oliver, B. (1960). Medicinal Plants in Nigeria. Published by Nigerian College of Science and Technology, Ibadan. Pp. 40.
6. Watt, J.M. and Breyer-Brandwijk, M.G. (1962). The Medicinal and Poisonous Plants of Southern and Eastern Africa. 2<sup>nd</sup> Edn. E and S Living Stone Ltd., London, pp. 687- 690.
7. Oketch – Rabah, H.A., Dosaji S.F., Christensen S.B. *et al.*, (1997). The antiprotozoal compound from *A. africanus*. *J. Nat. Prod.* **60**(10). 1017-1022.
8. Wall, M. E., Eddy, R.C, McClenman, M.L. and Klumpp, M.E. (1952). Detection and estimation of steroidal saponin in plant tissues. *Anal. Chem.* **24**: 1337.
9. Takeda, K. (1972): The steroidal saponins of the Dioscoreaceae. In: Progress in phytochemistry. Volume 2 (Edited by Reinhold, L. and Liwshitz, Y.) Interscience Publishers, London, pp. 287-33.
10. Faul, W. H. and Djerassi, C. (1970). Mass spectroscopy of Steroidal Saponins. *Organic mass spect.* **3**: 1187.